





### Determining the Impact of GIP Receptor Signaling on $\alpha$ -Cell GLP-1 Production

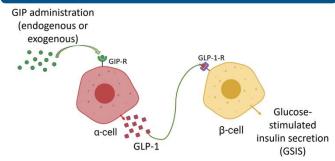
Catherine Dombroski<sup>1</sup>. Miranda Bustamante<sup>2</sup> & Bethany P. Cummings<sup>2</sup>

<sup>1</sup>University of California, Davis School of Veterinary Medicine, <sup>2</sup>University of California, Davis School of Medicine Department of Surgery

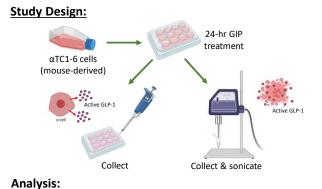
#### **Abstract**

Insulin secretion in response to oral glucose intake is described as the incretin effect and is driven by two gut-derived hormones: glucose-dependent insulinotropic polypeptide (GIP) and alucagon-like peptide-1 (GLP-1). GLP-1 is derived from the protein precursor proalucagon, which can be cleaved into either glucagon or GLP-1 via the PC2 or PC1/3 enzyme, respectively. Traditionally, it was believed that alpha cells express PC2 and not PC1/3, but multiple studies have demonstrated alpha cells can be stimulated to express PC1/3 and consequently produce GLP-1. Because the half-life of active GLP-1 in circulation is extremely short, it is hypothesized that alpha cells contribute to glucose-stimulated insulin secretion (GSIS) via paracrine signaling to beta cells using GLP-1. Indeed, a recent study demonstrated that GIP contributes to GSIS through the alpha cell. We hypothesize alpha cell GIP receptor signaling promotes GSIS by activating the production of GLP-1. The goal of our project is to determine if GLP-1 is released in response to alpha cell GIP receptor signaling, which would give more insight into potential mechanism behind alpha and beta cell communication in GSIS. To achieve this, we are treating alpha TC1-6 cells with GIP under conditions of high and low glucose and measuring the alpha cell response via ELISAs for active GLP-1 and alucagon levels. Additionally, PC1/3 & PC2 mRNA and protein levels will be quantified by aPCR and immunoblotting. Because glucogenic amino acids, such as alanine, are key stimulants of alpha cell hormone secretion, treatments of pancreatic islets with GIP and alanine will be analyzed as described above as well.

#### Hypothesis



#### Methods



## + + +

ELISA (for active GLP-1 & glucagon)

Western blotting (for PC1/3 & PC2)

# qPCR (for Pcsk1

#### & Pcsk2)

#### Results

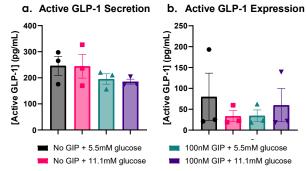


Figure 1. Active GLP-1 Expression and Secretion Following GIP Treatment of aTC1-6 Cells. (a) Active GLP-1 concentrations measured in conditioned media of  $\alpha$ -cells treated with GIP; n=3 per group (b) Active GLP-1 concentrations measured in Iysates of  $\alpha$ -cells treated with GIP; n=3 per group. Active GLP-1 levels quantified via sandwich electrochemiluminescence immunoassay (Meso Scale Discovery). Data are presented as means  $\pm$  SEM.

#### Discussion

- flux Secretion of active GLP-1 by lpha-cells does not change in response to GIP treatment under these experimental conditions.
- Expression levels of active GLP-1 also do not appear to be affected by GIP treatment under these experimental conditions.
- ☐ This experiment will be repeated to increase the sample size to determine any potential significance of our results.
- □ We repeated this study in human islets to assess the impact of GIP treatment on alpha cell GLP-1 production in a more translationally relevant model with intact paracrine signaling systems. These results are pending.

#### Conclusion

 $\Box$  The α-cell GIP-R plays a role in α/β-cell communication, but further investigation is required to identify the intermediate by which this occurs.

#### **Future Directions**

- Replicate experimental paradigm in human and mouse islets for increased translational and physiological relevance.
- ☐ Explore the role of glucogenic amino acids in glucosestimulated insulin secretion with GIP administration.

#### **Acknowledgements**

I would like to gratefully acknowledge my funding this summer from the NIH Grant T32GM136559 and UC Davis SVM Endowment Funds. This research project was made possible by funding from the Hartwell Foundation and NIH/NIDDK R56DK124853 grant. Figures made using BioRender.com

#### References

El, K., Gray, S. M., Capozzi, M. E., Knuth, E. R., Jin, E., Svendsen, B., Clifford, A., Brown, J. L., Encisco, S. E., Chazotte, B. M., Sloop, K. W., Nunez, D. J., Merrins, M. J., D'Alessio, D. A., & Campbell, J. E. (2021). GIP mediates the incretin effect and glucose tolerance by dual actions on a cells and β calls. Science Advances, 7(11). https://doi.org/10.116/j.cida/ab1f1948
Saikia, M., Holter, M. M., Donahue, L. R., Lee, I. S., Zheng, Q. C., Wise, J. L., Todero, J. E., Phuong, D. J., Garibay, D., Coch, R., Sloop, K. W., Garcia-Ocana, A., Danko, C. G., & Cummings, B. P. (2021). GIP-1 receptor signaling increases PCSK1 and β cell features in human α cells. J (In Sinth. 163). https://doi.org/10.1177/j.ci.nispit.14.14851

Timper, K., Dalmas, E., Dror, E., Rütti, S., Thienel, C., Sauter, N. S., Bouzakri, K., Bédat, B., Pattou, F., Kerr-Conte, J., Böni-Schnetzler, M., & Donath, M. V. (2016). Glucose-dependent insulinotropic peptide stimulates glucagon-like peptide 1 production by pancreatic islets via interleukin 6, produced by α cells. *Gastroenterology*, 151(1), 165–179. https://doi.org/10.1053/j.gastro.2016.03.003